

Themed Section: Science and Technology

Synthesis and Characterization of Six Co-Ordinate Complexes of Fe (III) Derived from N (4) Phenyl Thiosemicarbazone

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Fe (III) complexes have been synthesized by the reaction of FeCl₃with 5-chloro-2-hydroxy acetophenone phenyl thiosemicarbazone in presence of heterocyclic bases like pyridine (py), $\alpha/\beta/\gamma$ -picoline. Thiosemcarbazone has been characterized by ¹³C, ¹H NMR,ESI-MS as well as IR, electronic spectra. Spectral and magnetic measurements indicated Octahedral geometry for the six coordinate complexes. The metal complexes and their corresponding thiosemicarbazone were tested against bacterial parasites. It was found that the iron complexes synthesized are more biologically active then their corresponding thioemicarbazone.

Keywords: Metal Salts, Bioactivemetalcomplexes, Antimicrobialactivity, MIC.

I. INTRODUCTION

The complexes derived from salicylaldehyde and 2aminophenol were synthesised and characterised [1]. Magnetic electronic spectral studies provide the evidence of the existence of octahedral geometry for the complexes. Schiff base ligands by the reaction of salicylaldehyde and 2-hydroxy-1-naphthaldehyde 2-amino phenol with and 2-amino-p-cresol respectively have been prepared [2]. Co(II) complexes were studied by elemental analysis, 1H NMR, IR, UV-Visible spectra and TGA. The molar ratio of ligand to the metal of complexes is 1:1. Co (II) complexes have a hexa coordinated octahedral configuration. The redox process of ligands and complexes in DMSO solution were studied. Schiff bases derived from condensation of 2- hydroxy-1-naphthaldehyde and 2amino phenol have been studied [3]. The copper complexes were characterised by elemental analysis, molar conductance, electronic absorption spectra, IR and ESR spectral data and magnetic susceptibility data. The copper complexes as antibacterial agents were also studied.

The synthesis of new coordination compounds of Cu(II), Ni(II), Co(II), Sn(II), Hg(II) etc. with Schiff derived from 7-formyl-8-hydroxy bases quinoline(oxine) and 2-aminophenol have been reported by Sonbati and Bindary[4]. The ligands and the complexes were characterised by elemental analysis, IR, UV, EPR and NMR spectra together with magnetic susceptibility measurements. Infrared and NMR studies show that Schiff bases behave as monobasic and tridendate ligand, coordinating through the oxygen atom of the deprotonated phenolic group, the nitrogen atom of the azomethine group and pyridine.

The synthesis of mixed transition ligand metalcomplexes of Cu, Ni and Co ions with Schiff base ligands derived from the condensation of ohydroxy benzaldehyde with amino phenols and nitrogen donor amine bases, like ethylenediamine, 2o-phenylene aminopyridine, and diamine thiocyanate have been synthesised [5]. These characterised complexes were and their antibacterial, anti-fungal and toxicological activity have been evaluated.

In the present work the synthesis, spectral characterisation and biological assay of six coordinate complexes of Fe (III) with 5-chloro 2-hydroxy acetophenone N (4) thisemicarbazone have been reported.

II. EXPERIMENTAL

Materials and instrumentation

The thiosemicarbazone was synthesized by refluxing 5-chloro 2-hydroxy acetophenoneand N (4) phenyl thiosemicarbazide in ethanol in the mole ratio 1:1 for 3 hours. The pale yellow product obtained was filtered and washed with cold ethanol and then diethyl ether. It was recrystalised by hot ethanol and dried over P_2O_5 in vacuum.

Preparation of complex

FeCl₃salt was dissolved in absolute ethanol. Theethanolic solution of thiosemicarbazone in slight excess over the metal: ligand ratio of 1:1 was added to it dropwise and with constant

stirring. Therection mixture was refluxed for three hours. The dark brown product thus obtained was fitered and washed well with absolute ethanol and then diethyl ether and dried over P₂O₅ in vacuum.

$$\begin{array}{c|c} OH_2 \\ \hline OH_2 \\ \hline CI \\ \hline CH_3 \\ \hline CH_3 \\ \hline CH_3 \\ \hline OH_2 \\ \hline CH_3 \\ \hline OH_2 \\ \hline CH_3 \\ \hline OH_2 \\ \hline CH_5 \\ \hline CH_5 \\ \hline CH_5 \\ \hline OH_2 \\ \hline CH_5 \\ \hline OH_2 \\ \hline OH_3 \\ \hline OH_2 \\ \hline OH_2 \\ \hline OH_3 \\ \hline OH_3 \\ \hline OH_2 \\ \hline OH_3 \\ \hline OH_4 \\ \hline OH_5 \\ \hline OH_5$$

Preparation of complexes with heterocyclic bases

The complex Fe.L.B (B is heterocyclic base like pyridine, α -picoline, β -picoline, γ -picoline) was synthesized byadding slowly ehanolic solution of FeCl₃, heterocyclic baseto the hot ehanolic solution of

ligandin the ratio 1:1:1and refluxing reaction mixtureforthree hours. The brownadduct obtained was filtered and washed with hot water, cold ethanol and diethyl ether and dried over P_2O_5 in vacuum.

(B= pyridine, α -picoline, β -picoline, γ -picoline)

Physical measurements-

Magnetic susceptibility measurements were carried out by by Faraday method. IR spectra were recorded in the range 4000-200 cm⁻¹ range. NMR spectra were recorded in the mixture of CDCl₃ and DMSO-d₆ (1:1 v/v) with a Bruker AC-300F 300MHz spectrometer. Conductivity measurements were carried out on conductivity Bridge, Systonics conductivity meter-

304. Thermo gravimetric analysis was carried out in the temp rnge 30-800°C.UV-Visible spectra were measured on Jasco UV-visible double beam spectrophotometer.Metal in the complex and adducts was estimated by standerdizedxylenol orange as an indicator.Chloride in the complex was estimated by Mohr's method.

Table 1. Physical properties:

Compounds	Colour	Empirical Formula	Molar	Magnetic	
			conductance	Moment B.M.	
			Ohm ⁻¹ cm ² mole ⁻¹		
L	Yellow	C15H14ClN3OS	-	-	
Fe-L.Cl.(H ₂ O) ₂	Brown	C15H16N3O3Cl2SFe	40.0	5.20	
Fe.L.Py.(H ₂ O) ₂	Brown	C20H21N4O3ClSFe	43.0	5.25	
Fe.α-Pico.(H ₂ O) ₂	Brown	C21H23N4O3ClSFe	48.0	5.30	
Fe.L β-Pico.(H ₂ O) ₂	Brown	C21H23N4O3ClSFe	46.0	5.32	
Fe.L.y-Pico.(H ₂ O) ₂	Brown	C21H23N4O3ClSFe	48.0	5.27	

¹H-NMR

NMR signals at 13.00 and 3.1 ppm are assigned to – OH and – CH₃ protons respectively. Absence of $^2\mathrm{NH}$ proton signal suggests enolisation of $^2\mathrm{NH}$ – C = S group to $^2\mathrm{N=C-SH}$. Signal at 3.9 ppm cooresponds to $^4\mathrm{NH}$ Aromatic protons show multiples at 6.9, 7.20 , 7.60,7.65,7.77,7.30,6.20,7.29 ppm range.

¹³C-NMR (DMSO-D₆: δppm 122 (C=C), 135 (C=C), 130 (C=C-Cl), 130.78 (C=C), 125(C=C),165 (C=C-OH),170 (C=N),19 (=C-CH₃),187 (C=S),140 (NH-C=C),128 (C=C),130 (C=C),131 (C=C),130 (C=C),128 (C=C).

(Calcd) found ESI-MS m/z, ion M+:C15H14ClN3OS

319.18,C₁₅H₁₆N₃O₃Cl₂SFe (319.79)(445.26)

445.92,C₂₀H₂₁N₄O₃ClSFe

(488.91)

488.03,C₂₁H₂₃N₄O₃ClSFe

(502.93)

(502.93) 502.10,C₂₁H₂₃N₄O₃ClSFe

502.14,C₂₁H₂₃N₄O₃ClSFe (502.93) 502.11.

Analytical data

1. **L:** % C 56.07 (56.33),% H 4.87 (4.41),% N 13.80 (13.14),% S 10.89 (10.03)

2.**Fe-L.Cl.(H₂O)₂:** % Fe 13.01 (12.58),%Cl 7.09 (7.96),%C 40.02 (40.45),%H 3.07

(3.62),%N 9.97 (9.44),%S 7.80 (7.20).% O 10.78 (10.16) (6.37).% O 9.14 (9.54)

3. Fe.L.Py.(H₂O)₂:% Fe 11.90 (11.45),%C 49.78

(49.13),%H 4.91 (4.33),%N 11.97 (11.46),%S 6.01

(6.56).,% O 9.09 (9.82)

4.**Fe.α-Pico.(H₂O)**₂: % Fe 11.73 (11.13),%C 50.72 (50.15),%H 4.04 (4.61),%N 11.75 (11.14),%S 6.93

(6.37).% O 9.10 (9.54)

5.**Fe.L** β-**Pico.(H₂O)₂**: % Fe 11.70 (11.13),%C 50.70 (50.15),%H 4.10 (4.61),%N 11.70 (11.14),%S 6.90

(6.37).% O 9.12 (9.54)

6.**Fe.L.y-Pico.(H₂O)**₂: % Fe 11.77 (11.13),%C 50.78 (50.15),%H 4.12 (4.61),%N 11.78 (11.14),%S 6.90

Table 2. Electronic spectral data (cm⁻¹)

rable 2. Dieetronie spectrar data (cm.)							
Compound	Mode	MLCT	n→π*	π→π*			
L	DMF	-	30800	35806			
Fe-L.Cl.(H ₂ O) ₂	DMF	25700	32400	34700			
Fe.L.Py.(H ₂ O) ₂	DMF	25720	32430	34730			
Fe.α-Pico.(H ₂ O) ₂	DMF	25750	32450	34750			
Fe.L β-Pico.(H ₂ O) ₂	DMF	25786	32480	34770			
Fe.L.y-Pico.(H ₂ O) ₂	DMF	25725	32465	34790			

Infrared Spectroscopic data (cm⁻¹)

1.L: $v \leftarrow OH$) 3340; $v \leftarrow C = N$) 1678; $v \leftarrow C = S$) 790, 1375; v(N-N) 1075; $v(^2N-H)$ 3240; v(C-O) 1285.

2 [Fe-L.Cl.(H₂O)₂]:v (C = N) 1570; v (C = N-N=C) 1560, v (C-S) 710, 1301,v (N-N) 1125, v(M - N) 415, v (M-O) 525, v (M-S) 325, v (C - O) 1210,v(Fe-Cl) 370. $v(H_2O)$ 3530,3550.

3.[Fe.L.Py.(H₂O)₂]:v (C = N) 1575; v (C = N-N=C) 1565, v (C-S) 715, 1305; v (N-N) 1130, v (M - N) Base 250, v (M - N) 420, v (M - O) 530, v (M-S) 330, v (C - O)1215, Band due to HB 1450, v(H2O) 3540,3560. **4.**[Fe. α -Pico.(H₂O)₂]: ν (C = N) 1580; ν (C = N-N=C) 1570, v (C-S) 718, 1310,v (N-N) 1135, v (M - N) Base 255, v (M - N) 425, v (M - O) 535, v (M-S) 335, v (C -O) 1220, Band due to HB 1455, v(H₂O) 3545,3565. **5.**[Fe.L β -Pico.(H₂O)₂]: ν (C = N) 1585; ν (C = N-N=C) 1575, v (C-S) 720, 1315,v (N-N) 1140, v (M - N) Base 260, v (M - N) 430, v (M - O) 540, v (M-S) 340, v (C -O) 1230, Band due to HB 1460, v(H₂O) 3550,3570. **6.**[Fe.L.y-Pico.(H₂O)₂]:v (C = N) 1590; v (C = N-N=C) 1580, v (C-S) 725, 1320,v (N-N) 1145, v (M - N) Base 270, v (M - N) 435, v (M - O) 545, v (M-S) 345, v (C -O) 1240, Bands due to HB 1465, v(H2O) 3555,3575.

TGA analysis data:

The TGA curves of complexes were recorded between the temperatures 30 $^{\circ}\text{C}$ to 800 $^{\circ}\text{C}$

1.[Fe-L.Cl.(H₂O)₂]: First step, 114.0 °C, Mass loss 4.05 % second step, 134.0 °C, Mass loss, 8.50 % Third Step 240.0 °C, Mass loss, 32.0 % Fourth Step,

- 375.0 °C, Mass loss 63.0 %, Residue 800 °C, % of Fe₂O₃, 58.12 (58.39).
- 2.[Fe.L.Py.(H₂O)₂]:First step, 115 °C, Mass loss 3.20 % second step,130.0 °C, Mass loss, 7.10 % Third Step 250 °C, Mass loss, 35.0 %Fourth Step, 370.0 °C, Mass loss, 65.0 %, Residue, 790.0 °C, % of Fe₂O₃, 53.90 (53.18).
- 3.[Fe.α-Pico.(H₂O)₂]:First step, 115.0 °C, Mass loss 3.15 % second step, 130.0 °C, Mass loss, 7.76 % Third Step 249.0 °C, Mass loss, 30.0 % Fourth Step, 365.0 °C, Mass loss 62.0 %, Residue 800 °C, % of Fe₂O₃, 51.21 (51.70).
- 4.[Fe.L β-Pico.(H₂O)₂]:First step, 115.0 °C, Mass loss 3.17 % second step, 132.0 °C, Mass loss, 7.70 % Third Step 250 °C, Mass loss, 29.0 % Fourth Step, 366.0°C, Mass loss, 65.0 %, Residue, 779.0 °C, % of Fe₂O₃, 51.21 (51.70)..
- 5.[Fe.L.γ-Pico.(H₂O)₂]:First step, 114.0 °C, Mass loss 3.16 % second step, 131.0 °C, Mass loss, 7.77 % Third Step 245.0 °C, Mass loss, 33.0 % Fourth Step, 370.0 °C, Mass loss 60.0 %, Residue 800 °C, % of Fe₂O₃, 51.21 (51.70).

BIOLOGICAL ACTIVITY(AGAR PLATE DIFFUSION METHOD)

Table 3. Absorbances of solutions of L, Fe (III) complexes and standered

Compound	Staphyloco	Staphylococcu		Bacilus subtilis		Escherichia		Peudomonas	
	s aureu	s aureu				Co	li	aerugi	nosa
	Gram positive					Gram negative			
	2	2.5 μg/ml		2	2.5	2	2.5	2	2.5
	μg/ml			μg/ml	μg/ml	μg/ml	μg/ml	μg/ml	μg/ml
L	0.70	0.4	1	0.72	0.42	0.73	0.41	0.74	0.43
Fe-L.Cl.(H ₂ O) ₂	0.63	0.3	34	0.65	0.23	0.65	0.24	0.68	0.25
4Fe.L.Py.(H ₂ O) ₂	0.63	0.3	33	0.65	0.27	0.66	0.23	0.65	0.26
Fe.α-Pico.(H ₂ O) ₂	0.68	0.34		0.67	0.24	0.68	0.26	0.70	0.28
Fe.Lβ-ico.(H ₂ O) ₂	0.69	0.36		0.69	0.25	0.71	0.25	0.69	0.27
Fe.L.y-co.(H ₂ O) ₂	0.62	0.33		0.63	0.25	0.64	0.23	0.69	0.24
Fe-L.Cl.(H ₂ O) ₂	0.23	0.17		0.24	0.19	0.16	0.25	0.14	0.27
Standard	0.16	0.1	4	0.14	0.14	0.26	0.15	0.17	0.14

(Std-Amphiciline)

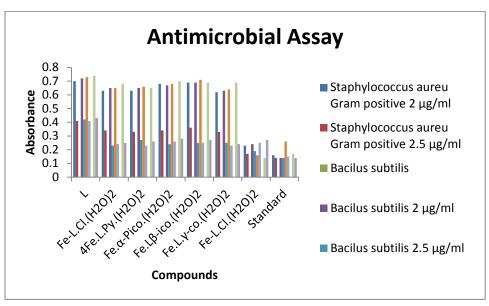


Figure 1. Antimicrobial Assay

III. RESULTS AND DISCUSSION

Physical data of thiosemicarbazone and its complexes are presented in Table 1. 1:1 ratio of metal ion, thiosemicarbazone for complex and 1:1:1 ratio for metal, thiosemicarbazone and heterocyclic base for all adducts was confirmed from elemental analysis. The solutions for conductivity measurement was made in and measurements were made 30°C.All complexes were found non electrolyte [6]. The magnetic moments were measured temperature 30°C .The magnetic moments are in between 5.20-5.32 B.M. Mass spectral data confirmed the structure of the thiosemicarbazone as indicated by molecular ion peak (M + 1) corresponding to their molecular weights.UV-visible spectral data presented in Table no.2 The thiosemicarbazone showed band π - π *band at 35,806 cm⁻¹ and n- $\pi^*[10,11]$ at 30,800 cm⁻¹[7-9]. The π - π^* absorption bands are shifted to longer wavelength in complexes due to the weakening of the C=S bond conjugation system gets enhanced on complexation [12,13]. The reduction in intensity of n- π *bands in the complexes is due to transfer of lone pair of electrons to the metal and coordination of azomethinenitrogen.MLCT bands are broad and observed in the region 25000-26000 cm⁻¹ which are assignable to Fe→S transitions.

Coordination through azomethine nitrogen shifts $^7C=N^1$ frequency to the lower side [14].New

bands in the range 415-435 cm⁻¹were observed confirms the participation of azomethine nitrogen in coordination [15,16]. There is increase in frequency of $^{1}N^{-2}N$. This confirms the coordination ofthiosemicarbazone to metal through the azomethine nitrogen atom. The band ²N-H of thiosemicarbazone disappears complexes in the indicates deprotonation of the ²N-H proton. A thioamide band partly due to C=S found at 1375 and 790 cm⁻¹ in thiosemicarbazone.It is shifted to lower side in complexes.This indicates coordination thiolate sulfur [17]. New bands in 325-340 range were oberved due to Fe-S confirms involvement of sulfur coordination. The phenolic oxygen indicates the third coordination on loss of OH protons. This shifts v(C-O)1285 cm⁻¹in thiosemicarbazone to lower side in the complexes. New bands observed due to Fe-O in complexes in the range 525-540 cm⁻¹ confirms coordination through oxygen. The heterocyclic base nitrogen atom occupies fourth coordination site. The band is assigned for v (Fe-N) due to heterocyclic base in 250-270 cm⁻¹ range in the spectra of all complexes. The characteristics bands due to coordinated heterocyclic bases are also observed in IR spectra of all complexes[18-20].

Themogravimetric analysis was carried out within a temperature range 30-800°C. The decomposition of complex proceeded in steps. Hydrated molecules of water in complexes were lost in between 30-110°C.

Then two coordinated water molecules were lost in the temperature range 114-116°C. Mass lost at this stage is about 4-5 % for one H₂O and upto 9 % for two H₂O molecules. No change observed ~ 130°C after that break observed in the curves due to evaporation of molecule of thiosemicarbazone. The remaining part of thiosemicarbazone removed from the coordination sphere at ~ 600°C. Finally the metal oxides were formed above 600°C. The complexes decomposed at ~ 800°C.

Fe (III) complex and adducts are thermally stable up to 114°C and decomposition started at this temperature. The temperature range for second step is 131-135°C. The third step temperature range is 245-250°C. The solid residue is Fe₂O₃. The complexes prepared with metal decomposed at higher temperature.. It is concluded that the thermal stability of the complexes is due to the coordination of metal ion to thiosemicarbazone [21].

The antibacterial assay was carried out by the agar plate diffusion method. Activity was measured by measuring the absorbance. Adducts prepared with showed good activity against heterocycic bases bacterial species than free thiosemicarbazone. The minimum inhibitory concentration was determined by liquid dilution method [22]. The solutions of thiosemicarbazone and complexes with 2 µg/ml,2.5 μg/ml and 3 μg/ml concentrations were prepared in the solvent DMF. The solutions of standard drug were also prepared in the same ampicilin concentration. Inoculums of the overnight culture were prepared.0.2 ml of the inoculums was added to the test tubes containing the solutions of the compounds of different concentrations. After the addition of sterile water to each of the test tubes, these were incubated for 24 hours and observed for turbidity. The absorbance of the turbid solutions was measured at 520 nm. The same method was used for standard [23].It was observed that at 2µg/ml the absorbance is more. Less absorbance at 2.5 µg/ml and no absorbance observed at 3µg/ml.The inhibition is more at 2.5 µg/ml. The inhibitory concentration is 2.5 µg/ml. Thus increase in coordination number and coordination of metal ion to ligand enhance microbial activity.

IV. REFERENCES

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